

SHORT COMMUNICATION

INFLUENCE OF UV LIGHT ON THE BIOSYNTHESIS  
OF ANTHOCYANIN-LIKE PIGMENTS IN  
RIPENING BANANAS

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(Received 29 July 1971)

**Abstract**—New compounds behaving like 3-desoxyanthocyanidin have been found to develop in banana fruit skin (*Musa sapientum*) as the result of UV exposure

INTRODUCTION

VISIBLE light is essential for the biosynthesis of anthocyanin in bean seedlings,<sup>1</sup> endosperm tissue culture of 'Black Mexican' corn,<sup>2</sup> *Spirodella oligorhyza*,<sup>3</sup> Milo seedlings,<sup>4</sup> strawberry leaf discs<sup>5</sup> and *Sinapis alba* plants<sup>6</sup> and UV light has been found to promote anthocyanin formation in *S. oligorhyza*.<sup>3</sup> However, most studies so far have been on plants which naturally form anthocyanin pigment. In banana, UV light<sup>8</sup> and  $\gamma$ -irradiation<sup>9</sup> has been reported to cause browning.

During studies concerning the storage of bananas (*Musa sapientum* L. var Dwarf Cavendish), an unusual red colouration was observed on the skin due to the effect of UV light exposure, in contrast to the yellow of normally ripe bananas. Since this red colour was similar to the red variety banana (*Musa coccinea*, locally known as 'Chandrabale'), the skin of which is known to have anthocyanins,<sup>10</sup> an attempt was made to characterize the pigments thus developed.

RESULTS AND DISCUSSION

The chromatographic behaviour, colour and spectral characteristics (as shown in Table I), together with the behaviour towards acid and alkali, of the pigments isolated from UV exposed Cavendish banana indicate that they are anthocyanins. The phenolic character of the pigments was clearly indicated by their reaction with Folin-Denis reagent and diazotized benzidine. Acid hydrolysis shows that the pigments are not glycosides. The  $R_f$ s and spectral characteristics, however, do not correspond to any known anthocyanidins.

<sup>1</sup> W H KLEIN, R B WITHROW, V ELSTAD and L PRICE, *Am J Bot* **44**, 15 (1957)

<sup>2</sup> J STRAUS, *Plant Physiol* **34**, 536 (1959)

<sup>3</sup> Y L NG, K V THIMANN and A G SOLON, *Plant Physiol* **36**, Suppl XLVI (1961)

<sup>4</sup> R J DOWNS and H W SIEGELMAN, *Plant Physiol* **38**, 25 (1963)

<sup>5</sup> L L CREASY, E C MAXIE and C O CHICHESTER, *Phytochem* **4**, 517 (1965)

<sup>6</sup> A HAVELANGE and R SCHUMAKER, *Bull Soc Roy. Sci Liege* **35**, 125 (1966)

<sup>7</sup> H MOHR, *Beitr Biochem Physiol Naturstoffen Festschr* 299 (1965)

<sup>8</sup> I HAUSTER, *Naturwissenschaften* **26**, 136 (1938)

<sup>9</sup> P THOMAS and P M NAIR, *Phytochem* **10**, 771 (1971)

<sup>10</sup> J B HARBORNE, *Chemistry and Biochemistry of Plant Pigments* (edited by T W GOODWIN), p. 273, Academic Press, New York (1965)

TABLE 1 CHROMATOGRAPHIC BEHAVIOUR AND SPECTRAL CHARACTERISTICS OF THE PIGMENTS

Pigment fraction	$R_f$ s in solvent		Absorption maxima in 0.01% HCl in methanol (nm)	Visual	Colour UV
	Formic acid	Forestal			
A	0.17	0.55	488	Orange	Pink absorption
B	0.22	0.64	533	Pink	Pink absorption
C	0.14	0.63	536	Light Pink	Strong orange fluorescence

*Note* Trace amount of fawn coloured compound having a higher  $R_f$  than these pigments was also noticed

On the basis of the  $R_f$ s, colour and the stability of these pigments, they appear to be 3-desoxyanthocyanidins (J B Harborne, private communication). No such anthocyanin has been reported in other parts of banana plant.<sup>11</sup> A similar red colouration was observed in 'Poovan' variety of banana, which was found to be proportional to the duration of the UV treatment.

The necessity of light for anthocyanin development is well established.<sup>1-7</sup> It appears that banana (*Musa sapientum*) has a dormant but weak anthocyanin forming system which, however, needs light of high energy for activation. UV irradiation has probably provided the required energy for the initiation and formation of this unusual anthocyanin.

#### EXPERIMENTAL

**Treatment** Green bananas (80–85 days after fruit set and maturity equivalent to the market samples with pulp to peel ratio being 1–1.3) were spread on a platform after 24 hr of separation from the bunch, and exposed to irradiation from a 30 W UV (200–280 nm lamp) from a distance of 45 cm for 40 min. The fruits were turned over after 20 min to maintain the uniform exposure.

**Extraction and purification of pigments** The outer thin skin of brownish red banana (about 15 days of UV irradiation, stored at 18–20°C), was separated from the peel and the pigments were extracted in cold in 0.1% methanolic HCl. The extract was initially purified by ascending paper chromatography using *n*-BuOH–2 N HCl. The different pigments were then separated by repeated paper chromatography using formic acid and Forestal solvents.

**Acknowledgements**—Grateful thanks are due to Dr J B Harborne, University of Reading for valuable help in the interpretation of results. Thanks are due to Dr A G Mathew and Mr J V Prabhakar for helpful discussions and to Mr H C Bhatnagar and Dr H A B Parpia, Director of this Institute, for their encouragement during the investigation.

<sup>11</sup> N W SIMMONDS, *Nature, Lond* **173**, 402 (1954)

<sup>12</sup> J B HARBORNE, *Biochem J* **70**, 22 (1958)

**Key Word Index**—*Musa sapientum*, Musaceae, 3-desoxyanthocyanidin, biosynthesis, UV light effect on pigment formation